

## A Study on Acinetobacter spp isolated from various clinical samples and analysis of their susceptibility pattern at a tertiary care centre

Wajid M.<sup>1\*</sup>, Gonti P.<sup>2</sup>, Mallamgunta S.<sup>3</sup>, Naaz S.<sup>4</sup>

DOI: <https://doi.org/10.17511/jopm.2021.i06.07>

<sup>1\*</sup> Mahamad Wajid, Associate Professor, Department of Microbiology, ESIC Medical College & Hospital, Sanath Nagar, Hyderabad, India.


<sup>2</sup> Prasanna Gonti, Senior Resident, , ESIC Medical College & Hospital, Sanath Nagar, Hyderabad, India.

<sup>3</sup> Saranya Mallamgunta, Assistant Professor, Department of Microbiology, ESIC Medical College & Hospital, Sanath Nagar, Hyderabad, India.

<sup>4</sup> Shazia Naaz, Assistant Professor, Department of Microbiology, ESIC Medical College & Hospital, Sanath Nagar, Hyderabad, India.

**Background:** Among the gram-negative bacterial infections, Acinetobacter spp infections are of critical importance given the severity of infections, intrinsic resistance to most antibiotics, and also capability to acquire new drug resistance. **Aim:** To study the prevalence of Acinetobacter spp and its antibiotic resistance patterns in a tertiary care hospital. **Materials and methods:** A retrospective study, including samples from various departments, submitted to the microbiology laboratory from January 2020 to December 2020. All the samples were processed according to standard conventional methods, and the Acinetobacter isolates were studied in detail about the demographic characters, speciation, and antibiotic susceptibility pattern. **Results:** Out of the 1242 positive isolates, 7.24 % were identified as Acinetobacter species; 90% were A. baumannii. Males were most common, most commonly belonging to 45-65 yrs. Out of all the recovered Acinetobacter spp, 30% were from ICU. 40% of the isolates were MDR, 74.07% from ICU and 26.98% from non-ICU settings were statistically significant ( $p < 0.005$ ). Cephalosporins showed the highest resistance to the isolates Ceftazidime 82.5%, followed by Gentamicin 73.3%. **Conclusion:** The present study showed that Acinetobacter spp prevalence and antibiotic resistance (MDR) are more common in ICUs. The emergence of carbapenem resistance in more than half the isolates is worrisome. The study suggests susceptibility testing to be done to help select the best antimicrobial drugs for therapy, thus helping in judicious use of antibiotics is necessary to prevent the emergence of antimicrobial resistance.

**Keywords:** Non-fermenters, ICU (Intensive care units), Drug resistance

Corresponding Author	How to Cite this Article	To Browse
Mahamad Wajid, Associate Professor, Department of Microbiology, ESIC Medical College & Hospital, Sanath Nagar, Hyderabad, India. Email: <a href="mailto:drmd.wajid@gmail.com">drmd.wajid@gmail.com</a>	Mahamad Wajid, Prasanna Gonti, Saranya Mallamgunta, Shazia Naaz, A Study on Acinetobacter spp isolated from various clinical samples and analysis of their susceptibility pattern at a tertiary care centre. Trop J Pathol Microbiol. 2021;7(6):313-319. Available From <a href="https://pathology.medresearch.in/index.php/jopm/article/view/588">https://pathology.medresearch.in/index.php/jopm/article/view/588</a>	

<b>Manuscript Received</b> 2021-11-17	<b>Review Round 1</b> 2021-11-19	<b>Review Round 2</b> 2021-11-26	<b>Review Round 3</b> 2021-12-03	<b>Accepted</b> 2021-12-10
<b>Conflict of Interest</b> Nil	<b>Funding</b> Nil	<b>Ethical Approval</b> Yes	<b>Plagiarism X-checker</b> 17%	<b>Note</b>



© 2021 by Mahamad Wajid, Prasanna Gonti, Saranya Mallamgunta, Shazia Naaz and Published by Siddharth Health Research and Social Welfare Society. This is an Open Access article licensed under a Creative Commons Attribution 4.0 International License <https://creativecommons.org/licenses/by/4.0/> unported [CC BY 4.0].



## Introduction

Gram-negative bacterial (GNB) infections are one of the most crucial health problems not only in the community but also in hospitalized patients. Due to the Lipopolysaccharide layer (LPS), GNB's, are known to cause sepsis at a higher rate and hence increased morbidity and mortality of patients [1].

Two large groups, Enterobacteriaceae and the non-fermenters, are responsible for most clinical isolates from cases of gram-negative infections [2].

Though the proportion of infection with non-fermenters is less when compared to that of Enterobacteriaceae, non-fermenters are of critical importance given the severity of infections they can cause and intrinsic resistance to most antibiotics [3].

Among the non-fermenters, the *Acinetobacter* and *Pseudomonas* are the two most important clinically significant pathogens. *Acinetobacter* spp is intrinsically resistant to several commonly used antibiotics, including aminopenicillins, first, second-generation cephalosporins and chloramphenicol [4].

Apart from the intrinsic property, they have a high capacity to acquire resistance to broad-spectrum  $\beta$ -lactams, aminoglycosides, fluoroquinolones and tetracyclines. They have emerged as a highly troublesome pathogen for many institutions especially in intensive care units (ICUs) globally, due to their immense ability to acquire or up-regulate antibiotic drug resistance determinants [5,6]. Their ubiquitous nature in the ICU environment and inadequate infection control practice has continuously raised.

There is an increase in the incidence of not only *Acinetobacter* infection over the past two decades [7] but also the emergence of MDR *Acinetobacter* and, in a few studies, even pan drug resistance. However, many studies are available giving the prevalence of *Acinetobacter* and also antimicrobial resistance patterns. Resistance rates can vary from person to person, geographic area and hospital setting. This study focuses on studying the prevalence of *Acinetobacter* spp isolated from various clinical specimens and also on identifying the portion of MDR *Acinetobacter* and comparison of their prevalence in ICU (Intensive care units) to Non-ICU (Non-Intensive care units) in a tertiary care hospital located in Hyderabad.

## Materials and methods

**Study setting:** The study was carried out in a 400 bedded hospital, tertiary care teaching hospital with 4 ICUs (medical, surgical, pediatric and neonatal), located in Hyderabad, Telangana.

**Type of study & methodology:** This is a retrospective study carried out over one year from January 2020 to December 2020. All the samples from various departments submitted to the microbiology laboratory for culture and antibiotic susceptibility during this one year were included in the study. All samples were subjected to routine microscopy, Gram staining and inoculated onto Blood agar and MacConkey agar for primary isolation and incubated aerobically at 37°C for 18-24 hours. Identification of isolates was performed by standard conventional methods based on the colony morphology, preliminaries like gram staining, catalase, oxidase, motility. Various biochemical tests were used to identify genus *Acinetobacter* like indole, citrate utilization test, urease test, triple sugar iron agar test, phenylalanine deaminase test. Identification of *Acinetobacter baumannii* species was made conventionally using specific tests like oxidative/fermentation glucose test, Arginine decarboxylation, and growth at 42°C [8]. Antibiotic susceptibility testing was performed by the Kirby Bauer disc diffusion method on Mueller-Hinton agar plates and interpreted according to the CLSI guidelines [9].

All the *Acinetobacter* isolates were tested for their antibiotic susceptibilities for various classes of antimicrobials using the following antibiotic discs: Cephalosporins (ceftazidime, ceftriaxone), Aminoglycosides, (Gentamicin, Amikacin), Fluoroquinolones (Levofloxacin, Ciprofloxacin), beta-lactam and beta-lactamase inhibitor combination drugs (Ampicillin + Sulbactam, Piperacillin + Tazobactam), carbapenems (imipenem, meropenem). Isolates showing resistance to three or more classes of antibiotics were categorized as (multidrug-resistant) MDR *Acinetobacter* spp. Isolates resistant to all commonly used antibiotics were pan-resistant [10].

**Inclusion criteria:** The study included all *Acinetobacter* species isolated from various specimens of all ages and different wards, including ICU and non-ICU settings.

**Exclusion criteria:** specimens with incomplete

Patient demographics, antimicrobial susceptibility testing reports that did not comply with Clinical and Laboratory Standards Institute guidelines (CLSI) were excluded.

All the Data was extracted from WHONET and was statistically analyzed using Epi-info version 7. Chi-square test and descriptive statistics were used to calculate the prevalence of *Acinetobacter* species and also the prevalence of MDR *Acinetobacter* in ICU and non-ICU settings. *P-values* less than 0.05 were considered statistically significant.

**Ethical consideration:** Institutional ethical committee approval was taken to conduct this study.

## Results

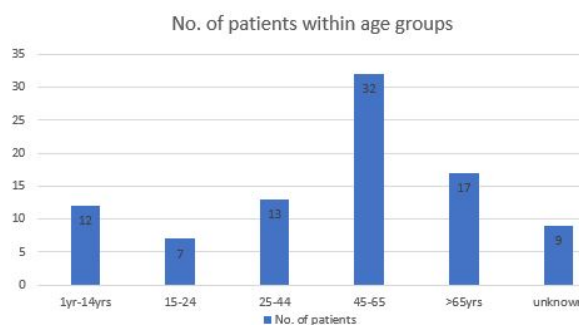
A total of 9,840 non-repetitive samples from various wards and ICU's [medical (MICU), surgical, pediatric, neonatal ICU's) were received in the Department of Microbiology, ESIC Medical College, Hyderabad, from January 2020 to December 2020, for culture and antibiotic susceptibility testing. Out of the 1242 positive isolates, 90 (7.24 %) were identified as *Acinetobacter* species. Among the 90 isolates, 81 (90%) were identified as *A. baumannii* and 9 (16.67%) as other *Acinetobacter* species. The demographic data of the specimens yielding growth of *Acinetobacter* spp showed males as the most common, with a male to female ratio of 2:1. Maximum number 32/90 of *Acinetobacter* isolates were recovered from 45-64 years age group (figure 1).

The distribution of various samples which yielded *Acinetobacter* isolates is discussed in fig 2, with endotracheal secretions 33% being the significant proportion. Other specimens are blood 25/90 (28 %), pus (24%), sputum (5) (6%), urine (3), and others 6%. (Graph 2).

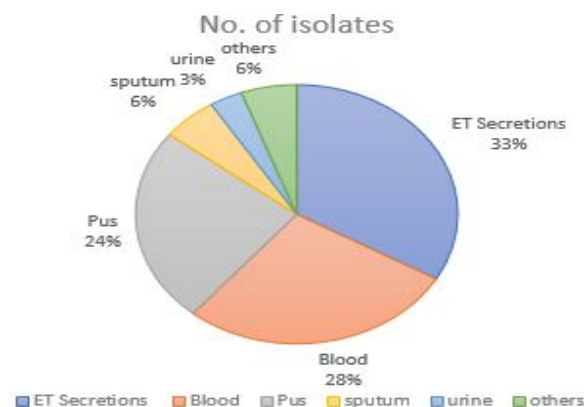
Out of all the *Acinetobacter* Spp that were recovered, 30% (27/90) were from ICU.

Antibiotic susceptibilities for the isolates are as per CLSI depicted in **figure 3**. Overall resistance percentages of *Acinetobacter* Spp for various classes of antibiotics the resistance patterns are as described. Cephalosporins had the highest resistance to the isolates (ceftazidime 82.5% is more resistant than ceftriaxone 78%), Aminoglycosides, (Gentamicin 73.3% slightly higher

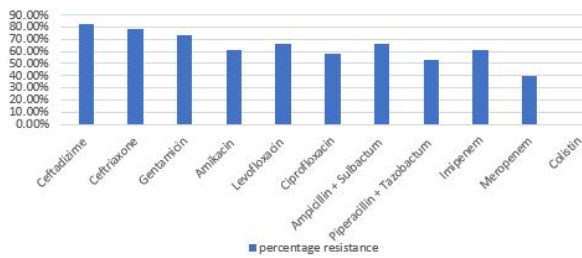
Resistance than Amikacin (61.4%) Fluoroquinolones (Levofloxacin 66.4% somewhat more resistant than Ciprofloxacin 57.9 %). When tested for beta-lactam and beta-lactamase inhibitor combination drugs, isolates showed slightly more resistance to Ampicillin + Sulbactam (66.7%) than Piperacillin + Tazobactam (52.8%). Nearly more than half of the isolates showed resistance to carbapenems. More number of isolates were resistant to imipenem (61.4%) as compared to meropenem (40%). Almost all the isolates showed resistance to any one of the classes of antibiotics mentioned above. Isolates showing resistance to three or more classes of antibiotics were categorized as MDR *Acinetobacter* spp. Isolates resistant to all commonly used classes of antibiotics except Polymyxin B were classified as pan-resistant. Out of 90 isolates, 36 (40%) were MDR. Because of MDR *Acinetobacter*, ICU isolates (74.07% %) and (26.98%) in non-ICU settings were MDR and were statistically significant (p <0.005). Among all the isolates, 6 were pan-drug resistance from ICU and one from non-ICU. 18.5% of isolates from ICU are found to pan drug-resistant. (Fig 4). Resistance to colistin was not identified in the present study.



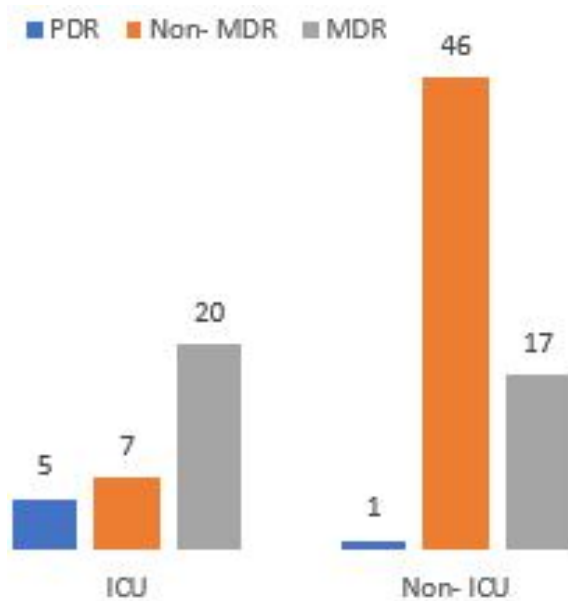
**Graph 1: Age-wise distribution of study population.**



**Graph 2: Percentage distribution of various clinical specimens which yielded Acinetobacter.**



**Graph 3: Resistance percentage of *Acinetobacter* spp to various antibiotics.**



**Graph 4: Distribution of MDR & PDR *Acinetobacter* among ICU and Non-ICU.**

## Discussion

*Acinetobacter* spp accounts for 7.24 % of all the positive cultures obtained from various specimens, similar to studies from Morocco [11] showing 6.94%, an international study compiling data of 75 countries [12]. showing (8.8%), studies from Rajasthan [13]. (6.42%) and Nepal [14] showing 11.4%, whereas studies from Punjab [15]. 9.6%. Among the 90 isolates, 81 (90%) were identified as *A. baumannii* and 9 (16.67%) as other *Acinetobacter* species similar to other studies where *Acinetobacter baumannii* was the most common species [16,17]. Kaur et al (91.6%) [15]. whereas W. Nageeb et al. [18] proved that *A.baumannii* was the only species isolated in clinical samples.

Out of the specimens that yielded *Acinetobacter*, 64.4% were from males, similar to studies by Uwingabaye et al. [11] and Ayenew Z et al. [19], showing 64.4 % & 60% respectively

And is in contrast to a study by Kaur et al. [15] where there is a female preponderance. The predominance of male patients infected with *Acinetobacter* has also been shown in other studies [20, 21]. The maximum number of isolates, 35.5%, were recovered from the 45-64 years age group, similar to a study by Uwingabaye et al. [11] with a similar age group of 42-68 years. In contrast, a study by Ayenew Z et al. [19] had a slightly younger population with a mean age of 30.79 as the most affected group. Samples that yielded *Acinetobacter* isolates were mostly 33% endotracheal secretions followed by blood 28%, similar to a study done by Uwingabaye et al. [11] and Rajesh et al. [13], where all the studies showed ET secretion as the major proportion. On the other hand, Kaur et al. [15] had shown urinary isolates to be the most common, in contrast to the present study. In the present study, 30% of the isolates were recovered from ICU patients. In contrast, a lower percentage, 11.4%, were isolated from ICU, as reported by Kaur et al. [15], and a higher percentage was reported by Rajesh et al. [13] Xia et al. [22]. and Tahseen and Talib [23]. The variation in these percentages could be due to the varying prevalence rates of *Acinetobacter* spp in different hospital settings and geographical areas. In the present study higher percentage of 82.5% of resistance was shown to the cephalosporin group of drugs which is similar to Kaur et al. [15]. and Saha S et al. [24]. About 73.3% of isolates showed resistance to aminoglycosides which is similar to studies by Raj kumari et al. [14]. 74.6%, and Taneja et al (72.8%) [25]. Levofloxacin is slightly more resistant than Ciprofloxacin 57.9 %, which is similar to Kaur et al. [15]. Though carbapenem is considered the treatment of choice, more than half of the isolates, 55.6%, were resistant to carbapenems in the present study, which is less when compared to studies showing (80%) by De Francesco MA et al. [26]. And (75%) by Shareek et al. [27]. The lower percentage could be due to the lesser proportion of patients from ICU in our study.

Out of 90 isolates, 36 (40%) were MDR similar to Dash et al. [28]. showing 54 %, De Francesco MA et al. [26] showing (54%), whereas studies from Kaur et al. [15]. 73.24%, Raj kumari et al [14]. 78. 6%, Saudi Arabia (74%) (29), from USA (30) 71. 6% and Bosnia and Herzegovina (78.4%) [19]. More isolates from ICU (74.07% %) were MDR than from non-ICU 26.98%. A similar trend

Of a higher percentage of MDR Acinetobacter in ICU was seen in a study by Uwingabiye J et al., showing 92.6% in ICUs and 75.3% and non ICU [11]. Among all the isolates, 6.6 % were pan-drug resistance, in the present study similar to Kaur et al. (8.5%) [15]. On the other hand, pan-resistant *A. baumannii* were not isolated in a study by Rajesh et al. [13]. Resistance to colistin was not identified in the present study, similar to Kaur et al. [15], Raj kumari et al. [14], Raina et al. [31], whereas Rajesh et al. [13]. identified (0.87%) colistin resistance.

## Conclusion

Various studies have indicated the role of Acinetobacter spp as a nosocomial pathogen, especially in ICUs, and have intrinsic resistance to many antibiotics apart from the capability to acquire them. Their prevalence varies depending on the geographical area and hospital setting. The present study has also shown that Acinetobacter spp is more common in the ICUs compared to the wards, and also the prevalence of MDR Acinetobacter spp. In the present study, Acinetobacter spp was most commonly resistant to cephalosporins and aminoglycosides. The emergence of carbapenem resistance in more than half the isolates is worrisome. In our study, colistin was the most sensitive antibiotic against Acinetobacter spp with a resistance rate of zero percent. The study suggests susceptibility testing to be done to help select the best antimicrobial drugs for therapy, according to the hospital setting. Also, judicious use of antibiotics is necessary to prevent antimicrobial resistance. Of course, not to forget the importance of adhering to hospital infection control practices both go hand in hand.

### What this study adds to the existing knowledge?

**Author's contribution:** **Wajid:** Supervision, correction of the manuscript, **Saranya:** data analysis, **Prasanna:** Collection of data. **Shazia:** Preparation of the manuscript.

## Reference

01. Ak O, Batirel A, Ozer S, Çolakoğlu S. Nosocomial infections and risk factors in the intensive care unit of a teaching and research hospital: a prospective cohort study. *Med Sci*

*Monit.* 2011 May;17(5):PH29-34. doi: 10.12659/msm.881750 [Crossref][PubMed][Google Scholar]

02. Oliveira, Junio, and Wanda C. Reygaert. Gram negative bacteria. (2019). [Crossref][PubMed][Google Scholar]

03. Perry JD. A Decade of Development of Chromogenic Culture Media for Clinical Microbiology in an Era of Molecular Diagnostics. *Clin Microbiol Rev.* 2017 Apr;30(2):449-479. doi: 10.1128/CMR.00097-16. Erratum in: *Clin Microbiol Rev.* 2017 Oct;30(4):vii [Crossref][PubMed][Google Scholar]

04. Seifert H, Baginski R, Schulze A, Pulverer G. Antimicrobial susceptibility of Acinetobacter species. *Antimicrob Agents Chemother.* 1993 Apr;37(4):750-3. doi: 10.1128/AAC.37.4.750 [Crossref][PubMed][Google Scholar]

05. Agodi A, Zarrilli R, Barchitta M, Anzaldi A, Di Popolo A, Mattaliano A, et al. Alert surveillance of intensive care unit-acquired Acinetobacter infections in a Sicilian hospital. *Clin Microbiol Infect.* 2006 Mar;12(3):241-7. doi: 10.1111/j.1469-0691.2005.01339.x [Crossref][PubMed][Google Scholar]

06. Goel N, Wattal C, Oberoi JK, Raveendran R, Datta S, Prasad KJ. Trend analysis of antimicrobial consumption and development of resistance in non-fermenters in a tertiary care hospital in Delhi, India. *J Antimicrob Chemother.* 2011 Jul;66(7):1625-30. doi: 10.1093/jac/dkr167 [Crossref][PubMed][Google Scholar]

07. Rungruanghiranya S, Somboonwit C, Kanchanapoom T. Acinetobacter infection in the intensive care unit. *J Infect Dis Antimicrob Agents.* 2005; 22(2): 77-9. [Crossref][PubMed][Google Scholar]

08. Collee JG, Fraser AG, Marmion BP, Simmons A. 14th ed. New York: Churchill-Livingstone; Mackie and McCartney Practical Medical Microbiology. 1999. [Crossref][PubMed][Google Scholar]

09. CLSI. Performance standards for antimicrobial disc susceptibility testing. 30th ed. CLSI supplement M-100 Wayne, PA: Clinical and Lab Standards Institute; 2020. [Crossref][PubMed][Google Scholar]

10. Dent LL, Marshall DR, Pratap S, Hulette RB.

- Multidrug resistant *Acinetobacter baumannii*: a descriptive study in a city hospital. *BMC Infect Dis*. 2010 Jul 7;10:196. doi: 10.1186/1471-2334-10-196 [Crossref][PubMed][Google Scholar]
11. Uwingabiye J, Lemnouer A, Roca I, Alouane T, Frikh M, Belefquih B, et al. Clonal diversity and detection of carbapenem resistance encoding genes among multidrug-resistant *Acinetobacter baumannii* isolates recovered from patients and environment in two intensive care units in a Moroccan hospital. *Antimicrob Resist Infect Control*. 2017 Sep 26;6:99. doi: 10.1186/s13756-017-0262-4 [Crossref][PubMed][Google Scholar]
12. Vincent JL, Rello J, Marshall J, Silva E, Anzueto A, Martin CD, et al. EPIC II Group of Investigators. International study of the prevalence and outcomes of infection in intensive care units. *JAMA*. 2009 Dec 2;302(21):2323-9. doi: 10.1001/jama.2009.1754 [Crossref][PubMed][Google Scholar]
13. Sannathimmappa, Mohan B. , Vinod Nambiar, and Rajeev Aravindakshan. Antibiotic resistance pattern of *Acinetobacter baumannii* strains: A retrospective study from Oman. " *Saudi Journal of Medicine & Medical Sciences* 9. 3 (2021): 254 [Crossref][PubMed][Google Scholar]
14. Rajkumari S, Pradhan S, Sharma D, Jha B. Prevalence and Antibiogram of *Acinetobacter* Species Isolated from Various Clinical Samples in a Tertiary Care Hospital. " *Journal of College of Medical Sciences-Nepal* 16. 1 (2020): 26-32. [Crossref][PubMed][Google Scholar]
15. Kaur R, Kaur S, Oberoi L, Singh K, Nagpal N, Kaur M. Prevalence & antimicrobial profile of *Acinetobacter* Spp. isolated from tertiary care hospital. *International Journal of Contemporary Medical Research* 2021;8(2):B1-B6. [Crossref][PubMed][Google Scholar]
16. Lone R, Shah A, Kadri SM, Lone S, Faisal S. Nosocomial multi-drug-resistant *Acinetobacter* infections-clinical findings, risk factors and demographic characteristics. " *Bangladesh Journal of Medical Microbiology* 3. 1 (2009): 34-38. [Crossref][PubMed][Google Scholar]
17. Joshi SG, Litake GM, Niphadkar KB, Ghole VS. Multidrug resistant *Acinetobacter baumannii* isolates from a teaching hospital. *J Infect Chemother*. 2003 Jun;9(2):187-90. doi: 10.1007/s10156-002-0224-4. PMID: 12872781 [Crossref][PubMed][Google Scholar]
18. Nageeb W, Kamel M, Zakaria S, Metwally L. Phenotypic characterization of *Acinetobacter baumannii* isolates from intensive care units at a tertiary-care hospital in Egypt. *East Mediterr Health J*. 2014 Apr 3;20(3):203-11. [Crossref][PubMed][Google Scholar]
19. Ayenew Z, Tigabu E, Syoum E, Ebrahim S, Assefa D, Tsige E. Multidrug resistance pattern of *Acinetobacter* species isolated from clinical specimens referred to the Ethiopian Public Health Institute: 2014 to 2018 trend analysis. *PLoS One*. 2021 Apr 29;16(4):e0250896. doi: 10.1371/journal.pone.0250896 [Crossref][PubMed][Google Scholar]
20. Punpanich W, Nithitamsakun N, Treeratweeraphong V, Suntarattiwong P. Risk factors for carbapenem non-susceptibility and mortality in *Acinetobacter baumannii* bacteremia in children. *Int J Infect Dis*. 2012 Nov;16(11):e811-5. doi: 10.1016/j.ijid.2012.07.006 [Crossref][PubMed][Google Scholar]
21. García-Garmendia JL, Ortiz-Leyba C, Garnacho-Montero J, Jiménez-Jiménez FJ, Pérez-Paredes C, Barrero-Almodóvar AE, et al. Risk factors for *Acinetobacter baumannii* nosocomial bacteremia in critically ill patients: a cohort study. *Clin Infect Dis*. 2001 Oct 1;33(7):939-46. doi: 10.1086/322584 [Crossref][PubMed][Google Scholar]
22. Xia Y, Lu C, Zhao J, Han G, Chen Y, Wang F, et al. A bronchofiberscopy-associated outbreak of multidrug-resistant *Acinetobacter baumannii* in an intensive care unit in Beijing, China. *BMC Infect Dis*. 2012 Dec 3;12:335. doi: 10.1186/1471-2334-12-335 [Crossref][PubMed][Google Scholar]
23. Tahseen U, Talib MT. *Acinetobacter* Infections As An Emerging Threat In Intensive Care Units. *J Ayub Med Coll Abbottabad*. 2015 Jan-Mar;27(1):113-6. [Crossref][PubMed][Google Scholar]
24. A study of *acinetobacter* infection in a tertiary care hospital in Northeast India. " *Int J Res Med Sci* 6 (2018): 2076-80. . [Crossref][PubMed][Google Scholar] [Crossref][PubMed][Google Scholar]
25. Taneja N, Singh G, Singh M, Sharma M. Emergence of tigecycline & colistin resistant *Acinetobacter baumannii* in patients with complicated urinary tract infections in north India. *Indian J Med Res*. 2011 Jun;133(6):681-4. [Crossref][PubMed][Google Scholar]

26. De Francesco MA, Ravizzola G, Peroni L, Bonfanti C, Manca N. Prevalence of multidrug-resistant Acinetobacter baumannii and Pseudomonas aeruginosa in an Italian hospital. *J Infect Public Health*. 2013 Jun;6(3):179-85. doi: 10.1016/j.jiph.2012.11.006 [Crossref][PubMed][Google Scholar]

27. Shareek, P. S. , et al. Antibiotic sensitivity pattern of blood isolates of Acinetobacter species in a tertiary care hospital: A retrospective analysis. " *American Journal of Infectious Diseases* 8.1 (2012): 65 [Crossref][PubMed][Google Scholar]

28. Dash M, Padhi S, Pattnaik S, Mohanty I, Misra P. Frequency, risk factors, and antibiogram of Acinetobacter species isolated from various clinical samples in a tertiary care hospital in Odisha, India. *Avicenna J Med*. 2013 Oct;3(4):97-102. doi: 10.4103/2231-0770.120501 [Crossref][PubMed][Google Scholar]

29. Almaghrabi MK, Joseph MRP, Assiry MM, Hamid ME. Multidrug-Resistant Acinetobacter baumannii: An Emerging Health Threat in Aseer Region, Kingdom of Saudi Arabia. *Can J Infect Dis Med Microbiol*. 2018 Feb 4;2018:9182747. doi: 10.1155/2018/9182747 [Crossref][PubMed][Google Scholar]

30. Rebic V, Masic N, Teskeredzic S, Aljicevic M, Abduzaimovic A, Rebic D. The Importance of Acinetobacter Species in the Hospital Environment. *Med Arch*. 2018 Nov;72(5):325-329. doi: 10.5455/medarh.2018.72.330-334 [Crossref][PubMed][Google Scholar]

31. Raina D, Sharma N, Mahawal BS, Khanduri A, Pandita A. Speciation and antibiotic resistance pattern of Acinetobacter species in a tertiary care hospital in Uttarakhand. " *International Journal of Medical Research & Health Sciences* 5. 4 (2016): 89-96. [Crossref][PubMed][Google Scholar]