

Study of coagulation profile in malaria


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Background: Malaria is a major health problem in many parts of India. Several factors have been attributed to increased morbidity and mortality in malaria with altered hematological and coagulation parameters. This study was conducted to compare the coagulation parameters of malaria cases with those of the healthy carriers. **Aims and objectives:** To evaluate coagulation abnormalities in patients of malaria and to study the difference in coagulation parameters between malaria patients and healthy controls and to determine the level of significance of the difference. **Materials and methodology:** This prospective comparative study of 300 patients with laboratory diagnosed malaria patients (cases) and 300 healthy individuals (controls) was carried out in the Department of Pathology in a tertiary care, V. S. General Hospital, Ahmedabad. **Result:** Comparison of platelet count, PT, and aPTT between case groups and control groups was statistically significant ($p < 0.001$). **Conclusion:** There is a significant difference between the platelet count, PT, and aPTT values of the two groups. This indicates that in patients with malaria, there is an activation of intrinsic and extrinsic pathways of coagulation.

Keywords: Malaria, Coagulation profile, Platelet count, PT, aPTT, D-dimer

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Introduction

Malaria is a mosquito-borne infectious disease caused by parasitic protozoans of the genus *Plasmodium* [1]. The parasites causing malaria are members of the Haemosporidia. These parasites of the genus *Plasmodium* are transmitted to human beings by a pre-infected female anopheles mosquito bite. Four species are generally recognized in man:

- (1) *P. falciparum* (the malignant tertian malaria parasite)
- (2) *P. vivax* (the benign tertian malaria parasite)
- (3) *P. ovale* (the benign tertian malaria parasite)
- (4) *P. malariae* (the quartan malaria parasite). *P. falciparum* produces the most serious form of the disease and is responsible for most malaria-related deaths [2].

A variety of hematological alterations like progressively increasing anemia, thrombocytopenia, leucopenia occur in malaria. The derangement in the coagulation profile in malaria is a highly sensitive measure to assess the severity and activity of the disease process. There is an accelerated coagulation cascade activity with accelerated fibrinogen turnover, consumption of antithrombin-III (AT-III), and increased concentration of fibrinogen degradation products.

Erythrocytes (RBCs) containing parasites are pro-coagulant. Released cytokines also act as pro-coagulants. Prothrombin time (PT) and activated partial PT (aPTT) is prolonged. Severe hemorrhage is reported in 5% of severe malaria [2]. The patient may develop bleeding gums, epistaxis, petechiae, subconjunctival hemorrhages, melena, and hematemesis [3].

Aims and Objectives

01. To evaluate coagulation abnormalities like-prothrombin time (PT), activated partial thromboplastin time (aPTT), and fibrinogen degradation product (D-dimer) in patients of malaria.
02. To study the difference in coagulation parameters between malaria patients and healthy controls and to determine the level of significance of the difference.
03. To understand the severity of coagulation defects and their related complications in patients with malaria.

Materials and Methodology

Type of study: Prospective study

Place and duration of study: This prospective comparative study was carried out in the Department of Pathology in tertiary care, V. S. General Hospital, Ahmedabad, and the duration of the study is 2 years.

Sample size: 300 patients with laboratory diagnosed malaria patients (cases) and 300 healthy individuals (controls)

Sampling methods: A comprehensive history including presenting complaints (Fever spikes, chills and rigor at regular intervals; classical malarial paroxysms: three stages- Cold stage, Hot stage and the Sweating stage and manifestations of severe *P. Falciparum* malaria), history, family history, drug history was taken by asking questions through a structured questionnaire. Patients of both sexes ranging from 3 years to 70 years were selected as per the below criteria:

Inclusion criteria:

- Cases: Indoor and outdoor malaria positive cases satisfying the following criteria were included in the study:
 - Age > 3 years of either sex
 - Slide positive malaria cases (*P. vivax* or *falciparum*)
 - Previously untreated for a present episode of
- Controls: Healthy individuals who did not have any features suggesting abnormalities related to hematological, biochemical, or coagulation parameters and not taking any anticoagulant therapy.

Exclusion criteria:

Patients with clinical history and or finding suggestive of

- Chronic liver disease
- Hemostatic abnormalities (aplastic anemia, leukemia, lymphoma, and vasculitis)
- Drugs used (sulphonamides, penicillin, cephalosporin, thiazide, and cytotoxic drugs).
- Congenital clotting factor defects (hemophilia, afibrinogenaemia, factor V and XI deficiencies)

Technical details:

Collection of blood:

01. For Hemoglobin, Total WBC count and Platelet count: 2 ml of blood was collected in EDTA vacuttee
02. For PT, aPTT, and D-dimer: 2 ml of venous blood was collected in 3.2% citrate

Processing of sample for coagulation studies:

Samples of the patients were immediately transported to the laboratory and were centrifuged at 3000-4000 revolutions for 15-30 minutes to obtain platelet-poor plasma (<10,000/cumm). Platelet poor plasma was used in PT, aPTT, and D-dimer. Samples were tested within 6 hours of collection of blood samples.

Investigations:

01. **Peripheral smear:** The clinical diagnosis of malaria was confirmed by peripheral blood smear examination which is the gold standard for the diagnosis of

For microscopic examination, peripheral blood thin and thick films were made on different slides (as per the method described in Practical Hematology-Dacie and Lewis) and stained with field’s stain. Grading of parasitemia (by semi-quantitative scale) was done from thick film examination.

02. **Hematological examination:** Complete hemogram with CELLDYN Ruby (Abbott) and Abbott CELLDYN 3700. Hemoglobin with less than 12.5 mg/dl was taken altered. Leucopenia with total white blood cell count <4000/cumm was taken altered. Thrombocytopenia with platelet count <1.5 lakh/cumm were taken

03. **Coagulation profile:** Was assessed by measuring prothrombin time (PT), activated partial thromboplastin time (aPTT), fibrinogen degradation products (D-dimer) by ACL Elite Pro Coagulation Analyser (Instrumentation Laboratory).

Statistical analysis:

- Comparison of platelet count, PT, and aPTT between case and control were done by using Mean±SD and students unpaired t-
- p-value <0.05 was taken to indicate a statistically significant
- All statistical analysis was done by using software Microsoft excel 2010 version and SPSS (statistical package for the social sciences) 21.0 version.

Results

Study shows that, out of 300 patients, 82 patients were affected by P. falciparum (27.3%), and 218 patients (72.7%) were affected by P. Vivax. Overall there were 7.67% pediatric patients in the study with P. falciparum patients being 10.9% of all P. falciparum cases and P. Vivax cases being 6.4% of all P. Vivax cases.

In both species, the majority of patients were adults. Out of 82 P. Falciparum cases, 26 (31.7%) were females and 56 (68.3%) were males. Out of 218 P. Vivax cases, 64 (29.3%) were females and 154 (70.7%) were males.

Table-1: Comparison of Mean between P. Falciparum and P. Vivax of baseline hemogram parameters.

		Number of cases	Mean Value	Minimum Value	Maximum Value
Hemoglobin	P. falciparum	82	9.4±2.8	4.2	16
	P. vivax	218	10.6±2.6	3.3	17.1
	Total	300	10.3±2.7	3.3	17.1
WBC Count	P. falciparum	82	7009.5±5107.3	1620	32700
	P. vivax	218	5875.9±2510.4	1390	19700
	Total	300	6185.8±3449.1	1390	32700
Platelet Count	P. falciparum	82	66258.3±45947.7	6000	227000
	P. vivax	218	98692.6±58810.6	8000	382000
	Total	300	89827.3±57373.6	6000	382000
RBC Count (lacs)	P. falciparum	82	3.7±1.1	1.5	5.9
	P. vivax	218	4.3±0.9	1.5	6.5
	Total	300	4.1±1.0	1.5	6.5

Anemia was observed in 55% of all patients, Leukopenia was observed in 22% of patients, and thrombocytopenia was seen in 85.7% of patients who presented with malaria.

Table-2: Changes in coagulation profile in malaria.

Parameter	Species		Total
	P. Falciparum	P. Vivax	
PT>15.3 Seconds	34 (41.4%)	95 (43.6%)	129 (43%)
APTT> 38 Seconds	26 (31.7%)	73 (33.5%)	99 (33%)
D-Dimer(>255ng/DI)	4 (4.9%)	1 (0.4%)	5 (1.7%)
Total Enrolled Patients	82 (100%)	218 (100%)	300 (100%)

PT was prolonged in 41.4% and 43.6% of P. falciparum and P. vivax patients respectively.

Similarly, APTT was prolonged in 31.7% and 33.5% of P. falciparum and P. vivax patients respectively.

D-Dimer was elevated in only 1.7% of all malaria patients.

Table-3: Comparison of coagulation profile between 300 patients with malaria and 300 healthy individuals (controls) were as follows.

Parameters	Study Group(n=300)	Control Group(n=300)	p- value (unpaired t-test)
PT	15.74±2.71	13.89±2.02	<0.001
Aptt	37.429±5.51	33.148±1.97	<0.001
Platelet Count	89827.267±57373.56	292427.93±50027.05	<0.001

Data was presented as Mean±SD, unpaired t-test was applied.

P-value of <0.05 was considered a statistically significant difference.

Thus, there was a statistically significant decrease in platelet count as well as an increase in PT and aPTT in patients with malaria as compared to controls.

Table-4: Comparison of coagulation profile between P. Falciparum and P. Vivax malaria were as follows.

Parameter	P. falciparum (n=82)	P. vivax (n=218)	p- value (unpaired t-test)
PT	15.56±2.85	15.81±2.66	0.5
aPTT	37.37±5.49	37.45±5.52	0.9
Platelet Count	66258.29±45947.67	98692.66±58810.59	<0.001

Data was presented as Mean±SD, unpaired t-test was applied. P-value of <0.05 was considered a statistically significant difference.

Thus, there was no significant difference in values of PT and aPTT amongst P. falciparum and P. vivax patients, however, platelet count was significantly lower in P. falciparum patients as compared to P. vivax patients.

Table-5: Comparison of Coagulation parameters in malaria among different studies.

Studies	Percentage of cases with altered Coagulation Profile		
	Altered PT	Altered aPTT	Altered Platelet
Present Study	43%	33%	85.7%
Previous studies	38%	56%	96%
Previous studies	34%	12%	68%
Previous studies	47.5%	35%	85%
Previous studies	21%	31%	63%

PT and aPTT were prolonged in 43% and 33% of the malaria patients respectively The derangement in values is comparable with those of Prasad R et al. and Kini et al who noted prolongation of PT in 47.5% and 38% of cases and deranged aPTT in 35% and 56% of cases, respectively.

Discussion

Pathophysiologic mechanisms of coagulopathy in malaria

1. Mechanisms of Thrombocytopenia in Acute Malaria

- Antibody-Mediated:

Antiphospholipid antibodies occur in patients with falciparum and vivax malaria and it has been suggested that they may be, at least partially, responsible for platelet activation and thrombocytopenia. Antiplatelet antibodies may activate platelet membranes, resulting in their removal by the hyperplastic reticuloendothelial (RE) system, particularly the spleen.

- Erythrocyte ADP Release:

It has been suggested that ADP, released by the hemolysis of erythrocytes activates platelets, which are then removed by the spleen.

- Platelet Phagocytosis:

Platelets are removed by activated macrophages in the spleen and liver.

▪ Oxidative Stress:

Oxidative stress may, through lipid peroxidation, cause premature platelet death, leading to malaria-associated thrombocytopenia.

▪ Platelet dysfunction:

During acute, *P. falciparum* and *P. vivax* infection, hyper aggregation and enhanced platelet secretory activity are demonstrated. This in vitro study suggested that the interaction between normal platelets and falciparum-infected erythrocytes could induce hypersensitivity of platelets, possibly through the stimulation of ADP released from infected red cells. Antibody bound to platelets as well as the invasion of platelets by malarial parasites may be other response mechanisms.

The other aspect of platelet dysfunction during malarial infection observed in some patients is the defective aggregation of platelets in response to ADP, epinephrine, and collagen and not ristocetin. From electron microscopic study, circulating degranulated platelets were observed during malarial infection. The presentation of the circulating exhausted platelets as a result of persistent in vivo activation is most likely a responsible mechanism causing the platelet hypoactivity [4].

2. Coagulation activation

During severe complicated malarial infection, the activation of the coagulation system leading to in vivo thrombin generation has been demonstrated. The stimulation of the coagulation system is caused by various procoagulants present during malarial infection. The sources of the procoagulants are exposed phosphatidylserine on the cell surface of infected erythrocytes, the lysis of activated platelets together with their secretory products, and the tissue factor (TF) released from damaged vascular endothelial cells. (4)

Furthermore, certain substances that are released during a severe malarial infection -namely tumor necrosis factor α (TNF α) and histamine - are additional factors that promote fibrin formation. The intrinsic pathway of the coagulation has also been shown to be activated in severe malaria. In turn, this may cause activation of the complement system and release of bradykinin and PMN-derived elastase that could contribute to the pathogenesis of severe malaria. Activation of the coagulation cascade also occurs in mild malaria.

The degree of activation is proportional to disease severity. Several sensitive indices of intravascular coagulation, including decreased plasma antithrombin(AT) activity and increased concentrations of thrombin antithrombin(TAT) complexes, are proportional to disease severity [5].

3. Defects in inhibitors of coagulation

Protein C, protein S, and AT levels were found to be low in *P. falciparum*, particularly in complicated cases, but were normal in *P. vivax* infection. The reduction in the levels of protein C, protein S, and AT is attributed to increased consumption due to microvascular thrombosis despite normal synthesis in the liver, as they correlated inversely with levels of TAT complexes [6].

▪ /Impaired fibrinolysis

Plasma levels of plasminogen activator inhibitor-1 (PAI-1) were very high in cases of *P. falciparum* infection as compared to normal controls and *P. vivax* infection. This could contribute to impaired fibrinolysis. The elevation of PAI-1 also strongly correlated with a decrease in the coma scale in cerebral malaria patients and correlated inversely with a reduction in platelet count, protein C, protein S, and AT levels. Both functional and antigenic tissue plasminogen activator (tPA) levels were low [6].

▪ Cytokines

Serum TNF α and interleukin-6 levels were elevated in the majority of patients with *P. falciparum* infection before antimalarial treatment. TNF α was also found to reduce the secretion of tPA and increase the secretion of PAI-1 [7].

Table-6: Acute versus “compensated” disseminated intravascular coagulation (DIC): an important distinction.

Profile of acute and compensated DIC		
Characteristic	DIC	
	Acute	Compensated
Prothrombin time	↑	N or S
Partial thromboplastin time	↑	N or S
Platelet count	↓	↓ or N
Fibrinogen level	↓	↓ or N or ↑
D- dimer	↑	↑ or N
Fibrinogen degradation products	↑	N or ↑
Fibrin monomer	↑	N or ↑
Thrombin-antithrombin complex	↑	↑
Plasmin-antiplasmin complex	↑	↑
Bleeding and hemorrhage	+	-

- Endothelial cell activation

The mechanisms involved in vascular endothelial cell damage in severe complicated malaria are multifactorial. A significant rise in plasma levels of both von Willebrand factor (vWF) and its propeptide, indices of chronic and acute endothelial cell perturbation, respectively. The increased levels indicate endothelial damage by the parasitized erythrocytes [7].

In uncomplicated and severe malaria, a coagulation disorder is a common laboratory finding, but bleeding and hemorrhage are observed in very few cases of severe malaria. Therefore, an important distinction should be made between these two pathologic states, one where typical DIC is encountered (e.g., bleeding), and the other where a compensated state (e.g., laboratory changes only) is detectable. DIC is characterized by activation of the coagulation cascade, which leads to the formation of microthrombi in the microcirculation, sometimes localized to a specific organ, but it often presents with an uneven distribution. Two major mechanisms trigger DIC: the release of TF into the circulation or endothelial injury.

While acute DIC is the terminal phase of the coagulation disorder, it is often preceded by a period during which the coagulation cascade is already activated but the increased activation can be compensated by the natural inhibitor systems, a state referred to as compensated DIC. The control mechanisms may effectively prevent severe clinical manifestations such as bleeding and hemorrhage by neutralizing active enzymes and/or by increasing the synthesis of the consumed hemostatic components.

As the trigger for coagulation activation persists in DIC, inhibitors will be gradually exhausted, leading to more coagulation. In this process, many clotting factors—like, fibrinogen and platelets—are consumed, resulting in eventually incomplete impairment of the hemostasis system. This is why the term 'consumptive coagulopathy' is often used to denote this process. This results in bleeding tendency or decompensated DIC.

Also, activation of the coagulation cascade is accompanied by compensatory fibrinolysis where an increase in plasminogen-dependent plasmin activity is detected using markers such as D-dimers, as shown in the above table. It also depicts the laboratory profile of acute and 'compensated' DIC [8].

Limitations

- The signs and symptoms of malaria may overlap with those of other infections. Therefore, the assessment of patients for other possible infections is of paramount importance.
- Leucocyte changes in malaria are variable and may not correlate exactly with the level of parasitemia.
- Thrombocytopenia may not be evident in some malaria cases, so careful peripheral smear examination is mandatory for accurate reporting.
- Derangement in coagulation profile may not be evident with the very low level of parasitemia.

Conclusion

There is a significant difference between the platelet count, PT, and APTT values of the two groups. This indicates that inpatient with malaria there is an activation of intrinsic and extrinsic pathways of coagulation.

What does the study add to the existing knowledge?

The present study concluded that activation of the coagulation cascade is not confined to *P falciparum* alone but is also seen in significant percentages of patients with *P vivax* infection. Moreover, most of the cases of malaria remain in a state of compensated DIC wherein the activated factors are kept in check by the anticoagulant mechanisms.

Author's contribution

All the authors, **Dr. Vibha Kantilal Patel, Dr. Nirali Kalpesh Shah, Dr. Anjali Deepak Goyal, Dr. Cherry Kalpesh Shah, Dr. Simul Jagdish Patel** have contributed to the concept, literature search, data acquisition, data analysis, manuscript editing, and review.

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