

# Prevalence and antibiotic susceptibility patterns of pseudomonas aeruginosa in urinary tract infections in a Tertiary care hospital, Central Kerala: A retrospective study over 4 years

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## Abstract

**Background and Objective:** Pseudomonas aeruginosa (P.aeruginosa) is an important uropathogen that has shown varied antibiotic susceptibility patterns. This study aims to find out the changing trends in the prevalence and antibiotic susceptibility patterns of urinary isolates of P.aeruginosa over four consecutive years. **Methodology:** A retrospective, record based study was conducted on all culture and sensitivity (C/S) reports of urine samples obtained in the microbiology lab in a tertiary care centre, Central Kerala (January 2014 -December 2017). The C/S reports which were positive for significant growth of P.aeruginosa were analyzed to find out its prevalence and antibiotic susceptibility patterns. Descriptive statistics were used for data analysis and the results were expressed in percentages. **Result:** Out of total 6622 urine samples received (14%) showed significant bacteriuria. P.aeruginosa was the third most common uropathogen isolated with an isolation rate of 3.5%. The antibiotic resistance observed were Gentamicin (53.1%), Amikacin (28%), Cefipime (28%), Ceftazidime (34.4%), Ciprofloxacin (43.7%), Norfloxacin (40.6%), Ofloxacin (40.6%), Piperacillin (37.5%), Piperacilli-Tazobactam (25%) and Imipenem (28%). The isolation rates of P.aeruginosa were 3.9%, 2.6%, 4.5 % and 2.9% in 2014, 2015, 2016 and 2017 respectively and over the years it maintained its third position. The year wise analysis of antibiotic resistance showed fluctuating pattern except Amikacin, Cefipime and Fluoroquinolones which displayed a decreasing trend. The reserve drugs like Piperacillin –tazobactam and Imipenem showed alarming drug resistance, although a hopeful reduction in the resistance was noted in 2017. **Conclusion:** P.aeruginosa remains as a common uropathogen. Drug resistant strains are markedly high in our area. Antibiotic resistance of P.aeruginosa does not show a consistent trend over years and vary from region to region. So each institution should have an antibiotic policy based on the local antibiogram which is to be renewed regularly. Instead of opting for higher antibiotic each time, strict implementation of restrictive and rotational antibiotic policies and adherence to the concept of ‘Reserve drugs’ should be followed. This is the only modality to inhibit the emergence of resistance strains of all uropathogens especially opportunistic pathogens like P.aeruginosa.

**Key words:** Antibiotic resistance patterns, Pseudomonas aeruginosa, Urinary tract infections.

## Introduction

Urinary tract infections (UTI) are the most common infections encountered by clinicians and one of the leading causes of morbidity in human population [1,2]. Although Escherichia coli is predominantly associated with the etiology of UTI, other organisms such as Klebsiella pneumoniae, Proteus mirabilis, Enterobacter, Citrobacter, Staphylococcus aureus, Enterococci etc account for most of the rest [1,3]. Aerobic non-

fermenting gram negative bacilli are now emerging as important uropathogens. Among these non-fermenters, P.aeruginosa is the predominant and most well-known organism [3,4].

P. aeruginosa is a ubiquitous, gram-negative bacillus that can survive in myriad of environment such as aquatic and terrestrial [4]. It is a versatile opportunistic pathogen, associated with nosocomial infections along with other serious implications with high rate of morbidity and mortality [5]. According to the report of

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**Original Research Article**

nosocomial infection surveillance system of center for disease control and prevention, *P. aeruginosa* is the third most common organism causing nosocomial urinary tract infections [5].

Treatment of UTI constitutes a great portion of prescription of antibiotics. Urinary pathogens have shown a changed pattern of susceptibility to antibiotics, showing an increased resistance to commonly used antibiotics due to extensive and inappropriate use of antimicrobial agents [6,7]. In *P. aeruginosa*, increasing resistance towards the available antimicrobials preclude the effectiveness of any antimicrobial regimen.

Because of increasing multidrug resistant (MDR) *P. aeruginosa* isolates in health care settings, infections are difficult to treat, causing life threatening conditions [5].

MDR mechanism in *P. aeruginosa* are due to lower outer membrane permeability, acquisition of enzymes like  $\beta$ -lactamases especially extended spectrum enzymes, carbapenemases or aminoglycoside modifying enzymes. Resistance is conferred by the transfer of plasmids which carry genes to produce antimicrobial enzymes [4,5].

Knowledge of the local bacterial etiology and susceptibility patterns are required to trace any change that might have occurred in time so that updated recommendation for optimal empirical therapy of UTI can be made. In this study we focused on *P. aeruginosa* because, this bacteria have different therapeutic options when compared to other commonly encountered gram negative uropathogens.

Even though a number of studies have been done on the prevalence and antimicrobial resistance patterns of uropathogens, no data have been reported from the present study area on *P. aeruginosa* causing urinary tract infections.

The aim of the study is therefore to determine changing trends in the prevalence of *P. aeruginosa* from suspected UTI cases and their antibiotic susceptibility patterns to the most commonly used antipseudomonal antibiotics.

**Result**

Out of 6622 urine samples received in our lab during the study period of 4 years, we got 925 (14%) culture positive cases with significant bacteriuria. The isolation rate of *P. aeruginosa* over the four years was 32 (3.5%). The year wise isolation rate of *P. aeruginosa* is shown in (Table:1).

The antibiotic susceptibility patterns of all *P. aeruginosa* isolates and year wise drug resistance pattern rates are shown in (Table: 2) and (Table:3) respectively.

**Materials and Methods**

A retrospective, record based study was conducted on all culture and sensitivity (C/S) reports of urine samples obtained in the microbiology lab at Sree Narayana Institute of Medical Sciences, Ernakulam during four consecutive years (January 2014 -December 2017).

**Inclusion criteria:** All Urine C/S reports with positive *P. aeruginosa* showing  $\geq 10^5$  colony forming units/ml.

**Exclusion criteria:** Urine C/S reports of *P. aeruginosa* from repeat culture of previously recruited patients.

**Method:** Mid stream urine samples were collected in sterile containers. The samples were cultured on blood agar and CLED agar (Cystine Lactose Electrolyte Deficient) medium with a standard loop and were incubated at 37°C overnight.

A growth of  $\geq 10^5$  colony forming units/ml was considered as significant bacteriuria. The *P. aeruginosa* isolates were identified by conventional biochemical test [8].

Antibiotic sensitivity testing was done by Kirby- Bauer disc diffusion method on Mueller-Hinton agar and interpretations were done according to the Clinical and Laboratory Standard Institute (CLSI) guidelines [9]. Antibiotics against which susceptibility tested were Gentamicin (10 $\mu$ g), Amikacin (30 $\mu$ g), Ciprofloxacin (5 $\mu$ g), Ofloxacin (5 $\mu$ g), Norfloxacin (10 $\mu$ g), Cefepime (30 $\mu$ g), Ceftazidime (30  $\mu$ g), Piperacillin (100  $\mu$ g), Piperazillin –Tazobactam (100/10  $\mu$ g), Imipenem (10 $\mu$ g).

Quality control was performed using *P. aeruginosa* ATCC 27853. The data regarding the number of *P. aeruginosa* and its susceptibility patterns were collected from the Microbiology lab register.

**Statistical analysis:** Descriptive statistics was used for analysis. Collected data were entered in MS-Excel and statistical analysis was done using SPSS 15 software and were expressed as percentages.

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**Table-1: Isolation rates of P.aeruginosa from urine samples.**

| Year         | Total number of Urine samples | Significant bacteriuria(%) | Isolation rate of P.aeruginosa (%) |
|--------------|-------------------------------|----------------------------|------------------------------------|
| 2014         | 1353                          | 230 (17)                   | 9 (3.9 )                           |
| 2015         | 1659                          | 235(14.2)                  | 6 (2.6)                            |
| 2016         | 1594                          | 222 (13.9)                 | 10 (4.5)                           |
| 2017         | 2016                          | 238 (11.8)                 | 7(2.9)                             |
| <b>Total</b> | <b>6622</b>                   | <b>925 (14)</b>            | <b>32 (3.5)</b>                    |

P.aeruginosa was the third most common urinary isolate after E.coli and Klebsiella species during the entire study period and year wise also it maintained its third position. The isolation rate was maximum in 2016.

**Table-2: Antibiotic resistance patterns of P.aeruginosa isolates (N=32)**

| Antimicrobial agents     | Number | Resistance (%) |
|--------------------------|--------|----------------|
| Gentamicin               | 17     | 53.1           |
| Amikacin                 | 9      | 28             |
| Cefipime                 | 9      | 28             |
| Ceftazidime              | 11     | 34.4           |
| Ciprofloxacin            | 14     | 43.7           |
| Norfloxacin              | 13     | 40.6           |
| Orfloxacin               | 13     | 40.6           |
| Piperacillin             | 12     | 37.5           |
| Piperacillin -tazobactam | 8      | 25             |
| Imipenem                 | 9      | 28             |

The drug resistance was highest against Gentamicin followed by Fluoroquinolones. Reserve drugs like Piperacillin – tazobactam and Imipenem exhibited similar resistance rates.

**Table-3: Year wise antibiotic resistance (%) of P.aeruginosa**

| Antimicrobial agents    | 2014 | 2015 | 2016 | 2017 |
|-------------------------|------|------|------|------|
| Gentamicin              | 44   | 66   | 50   | 57   |
| Amikacin                | 22   | 50   | 30   | 14.3 |
| Cefipime                | 33   | 50   | 20   | 14.3 |
| Ceftazidime             | 44   | 33   | 20   | 42.9 |
| Ciprofloxacin           | 44   | 83   | 40   | 14.3 |
| Norfloxacin             | 33   | 83   | 40   | 14.3 |
| Orfloxacin              | 33   | 83   | 40   | 14.3 |
| Piperacillin            | 44.4 | 66   | 20   | 28.6 |
| Piperacillin-tazobactam | 33.3 | 33   | 20   | 14.3 |
| Imipenem                | 44.4 | 50   | 10   | 14.3 |

Amikacin, Cefipime and Fluoroquinolones exhibit a decreasing trend over years but Gentamicin, Ceftazidime and Piperacillin shows an increasing pattern of drug resistance. When compared to 2015, Piperacillin- tazobactam and Imipenem shows a tremendous reduction in the drug resistance.

## Discussion

*P.aeruginosa* has established itself as a significant uropathogen which may cause dreaded complications if not treated properly. Antibiotic resistance is a major clinical problem in treating infections caused by this organism. The resistance patterns and isolation rates of *P.aeruginosa* varies regionally. Hence, increasing importance has been placed on the careful monitoring of antimicrobial resistance patterns of *P.aeruginosa* isolates for appropriate empirical as well as targeted treatment of the same. The study shows the prevalence and the antibiotic susceptibility patterns of urinary isolates of *P.aeruginosa* in a tertiary care hospital, Central Kerala.

In the present study *P.aeruginosa* was the third most common urinary isolate after *E.coli* and *Klebsiella* species. Other authors also observed *P.aeruginosa* as the third most frequent urinary isolate [10-13]. In some studies it acquired second position but in a recent study from Pakistan it ranked only fifth [2,4]. The isolation rates in the present study is compared to some recent studies from India and other countries (Table:4). Even though there is a slight variation in the prevalence, *P.aeruginosa* continue to be an important uropathogen in majorities of the studies.

**Table-4 : Prevalence of *P.aeruginosa* from urine samples in various recent studies**

| Various studies                        | Isolation rate of <i>P.aeruginosa</i> (%) |
|--|---|
| Present study                          | 3.5                                       |
| Bency JAT et al ; 2017; Kerala [10]    | 3.8                                       |
| Singh VP et al; 2017; UP [2]           | 6.7                                       |
| Sangeeta et al; 2017; Maharashtra [13] | 9.85                                      |
| Shah DA et al ;2015; Karachi [4]       | 5.4                                       |
| Jain et al;2014; Patiala [14]          | 9   |
| Prakash D et al; 2013; Meerut [12]     | 12.9                                      |
| Syed MA et al; 2012; Kerala [11]       | 2.74                                      |

The antibiotic resistance of *P.aeruginosa* is compared with various Indian and international studies in (Table:5).

**Table-5 : Comparison of Antibiotic resistance patterns of *P.aeruginosa* with various recent studies**

| Various studies          | Percentage of resistance (%) |          |          |             |               |             |           |              |                         |          |
|--------------------------|------------------------------|----------|----------|-------------|---------------|-------------|-----------|--------------|-------------------------|----------|
|                          | Gentamicin                   | Amikacin | Cefipime | Ceftazidime | Ciprofloxacin | Norfloxacin | Ofloxacin | Piperacillin | Piperazillin-Tazobactam | Imipenem |
| Present study            | 53.1                         | 28       | 28       | 34.4        | 43.7          | 40.6        | 40.6      | 37.5         | 25                      | 28       |
| Singh VP et al [2].      | -                            | -        | -        | -           | 20            | 14          | -         | -            | 10                      |          |
| Shah DA et al [4]        | 35.3                         | 25.3     | 63.9     | 56.1        | 50            | -           | 49        | -            | 19.6                    | 10.4     |
| Bency JAT et al [10]     | 30                           | 20       | -        | 90          | 90            | -           | -         | 30           | 10                      | -        |
| Syed MA et al [11]       | 49                           | 12.5     | 58       | 71          | 21            | 25          | -         | -            | 0                       | 0        |
| Sangeeta et al [13]      | 85.7                         | 85.7     | -        | 100         | -             | 42.85       | -         | -            | 71.4                    | 14.28    |
| Oladeinde BH et al [15]. | 100                          | -        | -        | -           | 40            | -           | 60        | -            | -                       | -        |
| Thomas ss et al [16]     | 47                           | 47       | 33       | 33          | 60            | -           | -         | -            | 33                      | 20       |
| Juayang C et al [17]     | 15.8                         | 6.7      | -        | -           | -             | -           | -         | -            | 9                       | 11.1     |

In the present study, 53.1% and 28% of the isolates were resistant to Gentamicin and Amikacin respectively. Over the 4 years, the resistance to Gentamicin was fluctuating but Amikacin showed a decreasing trend. The Aminoglycosides

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inhibit protein synthesis by binding to the 30 S subunit of the ribosome and the inactivation of the aminoglycosides occurs through the production of enzymes which transfer acetyl, phosphate or adenyl groups to the amino acid hydroxyl substituents on the antibiotics [16]. The resistance to Amikacin was highest during the year 2015 (50%), but it was lower than resistance to Gentamicin. This shows the growing resistance of *P.aeruginosa* to Gentamicin. Amikacin has been used sparingly only in severe forms of diseases owing to high cost and the intravenous nature of administration. Therefore, drug resistance has been slow to emerge. In our study, Amikacin is noted to be a comparatively effective drug. However, because of its numerous side effects including renal toxicity, blurred vision, hearing loss, Bartter-like syndromes, neuromuscular blockade, arthralgia, apnoea and many more, it is not commonly used [17].

Cefipime and Ceftazidime are the most frequently prescribed third and fourth generation Cephalosporins respectively [18]. In the present study, Ceftazidime showed 34.4% and Cefipime displayed 28% of resistance. A recent study from South India showed comparable resistance level but there are some studies showing a very high resistance to both [4,11,16]. In 2017 there was a tremendous decrease in the resistance against Cefipime (14.3%) but Ceftazidime resistance was almost thrice (42.9%) than that of Cefipime. The increased prevalence of Ceftazidime resistant *P.aeruginosa* can be related to inappropriate use of beta lactam antibiotics. Selective pressure from the use of antimicrobial agents is a major determinant for the emergence of resistant strains [18]. It is recommended to restrict use of Ceftazidime for a period of time to bring the developing resistance under control.

Among Fluoroquinolones, Ciprofloxacin, Norfloxacin and Ofloxacin showed almost similar resistance of 43.7%, 40.6% and 40.6% respectively. Different authors observed the resistance of ciprofloxacin as low as zero to as high as 90% [2,4,10,11,15,16,19]. Principal modes of Fluoroquinolone resistance in *P. aeruginosa* is due to target modifications in DNA gyrase (gyr A) and topoisomerase IV (par C) or mutations in regulatory genes for efflux pumps that reduce intracellular concentrations of the antibiotic [20]. Resistance for Fluoroquinolones decreased drastically and reached a promising low level of 14.3% in 2017. Similar study from Punjab also documented a decreasing trend over years [21]. This decreasing trend gives a promising evidence of rotational antibiotic policy in our institution.

25% of the isolates are Piperacillin –tazobactam resistant. Similar pattern of resistance to Piperacillin- tazobactam was observed by various authors but an extremely high resistance was noticed by a study conducted by Sangeeta et al [2,10,13,4,16,17]. Penicillins are highly ineffective against *P. aeruginosa* except for Piperacillin-tazobactam because of the beta lactamase inhibitor in addition to the extended spectrum and scarce use of the drug [4]. From 2014 to 2017, Piperacillin- tazobactam resistance remained almost consistent with a hopeful dip in 2017. Piperacillin –tazobactam continues to be a good choice of reserve drug for treating UTI caused by *P.aeruginosa*.

Resistance to Imipenem (28%) was also noted in this study. This is very high when compared to other recent studies [4,13,16,17]. This increased resistance is quite alarming, taking into account that Carbapenems are the last line of antibiotics for treating Gram-negative bacilli infections. Resistance to Carbapenems may be due to the result of complex interactions of several mechanisms including production of carbapenemase, over production of efflux system and loss of outer membrane porins. *P.aeruginosa* isolates that are Carbapenem resistant, specifically carbapenemase producing, are the worst, for the reason that they are associated with a higher mortality rate [17]. When compared to 2015, the resistance rate (50%), showed a tremendous decrease in 2017 (14.3%). This may be due to the strict implementation of antibiotic policy and infection control practices in our hospital.

**Conclusion**

*P.aeruginosa* remains as a common uropathogen. Drug resistant strains are markedly high in our area. The susceptibility pattern of one region differs widely from the other. The resistance of *P.aeruginosa* does not have a consistent trend over years. Irregular resistance pattern is observed except in some antibiotics such as Amikacin, Cefipime and Fluoroquinolones which showed a decreasing trend. Reserved drugs like Piperacillin- tazobactam and Imipenem showed an alarming drug resistance. It is emphasized that each

institution should have an antibiotic policy based on the antibiogram which should be renewed yearly. Instead of going for higher options of antibiotic each time, strict implementation of restrictive and rotational antibiotic policies and adherence to the concept of ‘Reserve drugs’ should be followed by each institution.

This is the only modality to inhibit the emergence of resistance strains of all uropathogens especially opportunistic pathogens like *P.aeruginosa*.

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**What's New in this study:** This study provides information regarding the prevalence and anti microbial susceptibility pattern of urinary isolates of *P.aeruginosa*.

The study stresses on the importance of C/S reports provided by the microbiology laboratory, so that clinician can select the appropriate antibiotic therapy.

Itemphasizesthe importance of close monitoring of antibiotic susceptibility patterns by preparation of antibiogram and its regular updating.

The study also intends to motivate the strict implementation of restrictive and rotational antibiotic policies and adherence to the concept of reserve drugs.

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